



Review Article

Association of fetuin-A with the pathogenesis of metabolic diseases



Open Access

Reza Afrisham¹, Seyyed Mohammad Reza Hashemnia¹, Ziba Majidi¹, Sadegh Mozaffari¹, Mahmoud Vahidi*²

ARTICLE INFO

Article History:

Received 8 March 2020

Revised 28 March 2020

Accepted 18 June 2020

Keywords:

Fetuin-A

Obesity

Diabetes

Non-alcoholic fatty liver disease

Cancer

¹Department of Clinical Biochemistry, Faculty of Medicine, Tehran University of Medical Sciences, Tehran, Iran;

²Department of Laboratory Sciences, Faculty of Paramedicine, AJA University of Medical Sciences, Tehran, Iran.

Correspondence:

Mahmoud Vahidi, Department of Laboratory Sciences, Faculty of Paramedicine, AJA University of Medical Sciences, Tehran, Iran.

Email: mahmoud.vahidi@gmail.com

ABSTRACT

Introduction: Fetuin-A is a glycoprotein that is synthesized by liver cells. Studies have shown that this hepatokine is linked to various metabolic disturbances such as obesity, and diabetes. Therefore, this study was designed to evaluate the role of fetuin-A in the pathogenesis of metabolic disturbances.

Methods: The present study was a review article. The terms of “fetuin-A”, “diabetes”, “obesity”, “chronic kidney disease (CKD)”, “cardiovascular disease (CVD)”, “non-alcoholic fatty liver disease (NAFLD)”, “cancer”, “bone metabolic diseases” and “metabolic disease” were used for searching of research papers in databases including Embase, Scopus, Web of Science, PubMed and Google Scholar. The literature search was limited to papers published up to November 2019.

Results: Fetuin-A could be involved in the pathogenesis of metabolic diseases such as obesity, diabetes, CKD, CVD, cancer, bone metabolic diseases, and NAFLD through various signaling pathways.

Conclusion: The results of the current study showed that fetuin-A could be involved in the pathogenesis of the metabolic disease. However, the study on these findings needs further research and a better understanding of these pathomechanism communications, which can be promising and helpful in preventing and better targeting metabolic disorders.

Introduction

Analogous to the myokines and adipokines, the hepatic proteins are named hepatokines (1, 2). Fetuin-A is a 64 kDa glycoprotein that is found in relatively high concentrations (100-1000 µg / ml) in human serum (3). This glycoprotein is also known in humans as alpha2-Heremans-Schmid glycoprotein (AHSG) (4). Fetuin-A is secreted from hepatocytes and is considered as hepatokine (4). Until recently, it was believed that the liver was the only major organ that secretes fetuin-A, but later it was found that adipocytes could also synthesize fetuin-A (5). The highest values of this hepatokine are observed in fetal blood that is synthesized in different tissues, accordingly, the name fetuin is applied (6). Epidemiological studies have consistently shown that increased levels of fetuin-A can be seen in obesity and obesity-related complications, such as type 2 diabetes mellitus (T2DM) (3), metabolic syndrome, and non-alcoholic fatty liver disease (NAFLD) (7).

Studies have also indicated that fetuin-A concentration was positively correlated to leptin concentration, visceral adipose tissue, and body mass index (BMI) (7). Moreover, circulating fetuin-A is higher in insulin-resistant obesity than insulin-sensitive obesity (7) so that, fetuin-A interferes with insulin signaling at the level of tyrosine kinase receptor (8) and lead to a

higher risk of T2DM (3). Also, it has been established that the single nucleotide polymorphism (SNP) of the AHSG gene is associated with T2DM (7). Also, it has been reported that there is a positive correlation between fetuin-A levels and stage of liver fibrosis (7, 9). In cancers, in addition to the liver, cancer cells themselves also synthesize and release fetuin-A (10-12). Studies show that fetuin-A plays a role in enhancing cancer cell attachment (13), motility, and invasion (14, 15). Given the changes of fetuin-A in metabolic diseases; we aimed to evaluate the molecular and cellular mechanisms linking hepatokine fetuin-A to the pathogenesis of metabolic diseases.

Methods

The present study was a review article. The terms “fetuin-A”, “diabetes”, “obesity”, “chronic kidney disease”, “cardiovascular disease”, “non-alcoholic fatty liver disease”, “cancer”, “bone metabolic diseases” and “metabolic disease” were used for searching research papers in databases including Embase, Scopus, Web of Science, PubMed and Google Scholar. The literature search was limited to papers published up to November 2019.

Results

Association of fetuin-a with obesity

Obesity is a metabolic disorder that is considered as one of the important risk factors for CVD, NAFLD, and T2DM (6). Adipocytes in addition to hepatocytes could synthesize fetuin-A (5). Experimental and clinical studies have shown that fetuin-A has an obvious role in obesity (16). In 2017, Pérez-Sotelo et al. demonstrated that visceral adipose tissue more than subcutaneous adipose tissue secreted fetuin A that was sensitive to physiological and nutritional changes. They suggested that adipose-fetuin A played a key role in the deleterious effects of obesity (17). It has been established that leptin concentration, visceral adipose tissue, and BMI are positively correlated with serum fetuin-A concentrations (7). Positive energy balance in mice has been shown to increase fetuin-A mRNA expression levels (18). Besides, it has recently been shown that a positive energy balance in humans increases blood fetuin-A levels (19). After gastric bypass surgery along with a dramatic weight loss, a significant reduction in the levels of this hepatokine was observed (20).

It was very interesting that Mathews et al. indicated that when fetuin-A knockout mice fed a high-fat diet, a significant reduction of body fat, as well as resistance to the weight gain, was observed as compared to controls (21). In 2010, Dasgupta et al. showed that the incubation of HepG2 cells or mouse hepatocytes with palmitate stimulated NF- κ B (nuclear factor- κ B) binding to the fetuin-A promoter; leading to increased levels of mRNA expression, protein

synthesis, and the secretion of fetuin-A (22). Jung et al. evaluated the influences and regulatory mechanisms of adiponectin and salsalate on fetuin-A expression in palmitate-induced HepG2 cells. Palmitate elevated SREBP-1c and fetuin-A expression and resulted in steatosis, while, knockdown of this hepatokine restored these changes. Adiponectin remarkably suppressed palmitate-stimulated fetuin-A mRNA expression and secretion through the adenosine monophosphate-activated protein kinase (AMPK) pathway. They concluded that adiponectin and salsalate improved impairment of lipid metabolism and palmitate-induced steatosis in HepG2 cells by the inhibition of this hepatokine through the AMPK-NF κ B pathway (23). Thus, the decreased adiponectin, which is often seen in obesity (24), maybe another reason for the increase of fetuin-A in obese individuals (7). On the other hand, fetuin-A exacerbates this pathway and downregulates this adipokine in lipid-induced inflamed adipocytes (6).

Moreover, fatty acids stimulate fetuin A protein and gene expressions in adipocytes, leading to its copious release. Chatterjee et al. in 2013, showed that lipid-induced fetuin A from adipocytes acts as a chemoattractant factor for migration and polarization of macrophages which polarizes M2 macrophages to M1 (pro-inflammatory macrophages) (25). This hepatokine also induces secretion of pro-inflammatory cytokines such as IL-6 and TNF-alpha in adipose tissue (26). This inflammatory condition can contribute to the initiation and progression of other metabolic diseases, including insulin resistance, T2DM, NAFLD, CVD, and even cancer. Table 1 shows the association of fetuin-A with various metabolic diseases.

Table 1. Association of fetuin-A with various metabolic diseases

Studies	The metabolic disease	The involved tissue/cells/animal	Results
Pérez-Sotelo et al. (17)	Obesity	Adipose tissue	Visceral adipose tissue more than subcutaneous adipose tissue secreted fetuin A. This hepatokine played a key role in the deleterious effects of obesity
Mathews et al. (21)	Obesity	Fetuin-A knockout mice	Fetuin-A knockout mice fed a high-fat diet, a significant reduction of body fat as well as resistance to weight gain observed as compared to controls
Dasgupta et al. (22)	Obesity/NAFLD	Liver tissue	The incubation of HepG2 cells or mouse hepatocytes with palmitate stimulated NF- κ B (nuclear factor- κ B) binding to the fetuin-A promoter; leading to increased levels of mRNA expression, protein synthesis, and the secretion of fetuin-A
Jung et al. (23)	Obesity//NAFLD	Palmitate-induced HepG2	Adiponectin and salsalate improved impairment of lipid metabolism and palmitate-induced steatosis in HepG2 cells by the inhibition of this hepatokine through the AMPK-NF κ B pathway
Chatterjee et al. (25)	Obesity	Immune cells	Lipid-induced fetuin-A from adipocytes acts as a chemoattractant factor for migration and polarization of macrophages which polarizes M2 macrophages to M1
Goustin et al. (31)	Diabetes	Mouse muscle cells	Human fetuin-A interferes with insulin receptor signaling at the level of tyrosine kinase. Fetuin-A inhibits insulin-stimulated GLUT4 translocation and activation of protein kinase B in mouse muscle cells and interferes with downstream phosphorylation of the insulin signaling pathway without affecting insulin binding to the alpha receptor subunit.
Zhao et al. (46)	Cardiovascular Disease	Serum	Subjects without CAD had significantly lower levels of serum fetuin-A
Akin et al. (47)	Cardiovascular Disease	Serum	Fetuin-A was significantly elevated in patients with CAD in comparison with patients without CAD
Chen et al. (34)	Cardiovascular Disease	Serum	Lower level of fetuin-A was related to an elevated risk of CVD and all-cause mortality in CAD patients.
Bilgicir et al. (40)	Cardiovascular Disease	Serum	Fetuin-A was lower in myocardial infarction and stable angina patients as compared to control group
Naito et al. (49)	Cardiovascular Disease	Human umbilical vein endothelial cells <i>in vitro</i>	Fetuin-A induced the expression of intercellular adhesion molecule-1 (IAM-1), monocyte chemoattractant protein-1 (MCP-1), E-selectin, and IL-6
Yusuf Yilmaz and et al. (9)	NAFLD	Serum	Serum level of Fetuin-A significantly increases in adult patients with NAFLD compared to healthy controls and a significant correlation was observed between serum Fetuin-A and liver fibrosis score index
Zhengsen Cui et al. (63)	NAFLD	Serum	fetuin-A in patients with NAFLD decreased when compared to healthy ones
Guillory et al. (76)	Cancer	Mouse	Lacking of fetuin-A reduces breast tumor incidence, extends latency, and decreases attenuation of aging caused by oncogenes. These events were triggered by an increased TGF- β signaling pathway and decreased phosphatidylinositol 3-kinase/Akt signaling pathway
Babler et al. (102)	Metabolic Bone Diseases	Mouse	The calcified soft tissues of mice lacking fetuin-A, after exposure to this glycoprotein, it returns to normal
Sari et al. (107)	Metabolic Bone Diseases	Serum	Fetuin-A levels are lower in postmenopausal osteoporotic patients than in controls
Caglar et al. (114)	Chronic Kidney Disease	Serum	In a group of nondiabetic subjects with different stages of CKD, fetuin-A levels diminished with a reduction in glomerular filtration rate
Coen et al. (115)	Chronic Kidney Disease	Serum	Hemodialysis patients with lowest tertile of fetuin-A had the maximum CRP levels
Mehrotra et al. (110)	Chronic Kidney Disease	Serum	The association between high levels of serum fetuin-A and decreased renal clearance and remarkable proteinuria in diabetic subjects

Association of fetuin-A with diabetes

T2DM is a chronic metabolic disorder that results from insulin resistance (27) and is considered as one of the important risk factors for CVD, and NAFLD (6). As mentioned above, fetuin-A has an obvious role in obesity (16). On the other hand, it has been reported that circulating fetuin-A is higher in insulin-resistant obesity than insulin-sensitive obesity (7). Also, it has shown that the risk of diabetes is higher in women and/or men with elevated concentrations of fetuin-A (3). Studies have shown subjects with T2DM had higher levels of hepatokine in comparison with nondiabetic individuals (26, 28, 29). Genetic studies observed that the SNP of the AHSG gene is associated with T2DM (7) as well as both known susceptibility loci for T2DM and the gene encoding this hepatokine are localized on chromosome 3q27 (30). In addition to this study, many studies have also shown that elevated fetuin-A levels can cause insulin resistance in liver cells (6, 8).

Some possible mechanisms might describe the relation between fetuin-A and T2DM. This glycoprotein is an endogenous inhibitor of the insulin receptor tyrosine kinase in muscle, fat, and liver tissue (8). In another study in 2013, Goustin et al. showed that human fetuin-A interferes with insulin receptor signaling at the level of tyrosine kinase. In fact, fetuin-A inhibits insulin-stimulated GLUT4 translocation and activation of protein kinase B in mouse muscle cells. This hepatokine also interferes with downstream phosphorylation of the insulin signaling pathway without affecting insulin binding to the alpha receptor subunit (31). Besides the direct effects of fetuin-A on the insulin receptor, it can also increase insulin resistance by inducing an inflammatory state. So that, treatment with fetuin-A increases the mRNA and protein expression of proinflammatory cytokines in both adipocytes and monocytes (7, 22). A recent study reported that fetuin-A is essential for non-esterified fatty acids to induce inflammation and insulin resistance through signaling Toll-like receptors (TLR4) in both adipocytes and macrophages (32). On the other hand, adiponectin is an adipokine with insulin-sensitizing and anti-inflammatory function that is reduced by fetuin-A (33) and the decreased adiponectin may be another mechanism for the fetuin-A induced insulin resistance.

Association of fetuin-A with cardiovascular disease

Obesity, T2DM, and NAFLD are the major risk factors for CVD (6). cardiovascular disorders are considered the leading reason for mortality worldwide (34). Fetuin-A serves a dual function in cardiovascular disorders (35), so that; a) high levels of this factor are correlated to metabolic disorders related to CVD including obesity, T2DM, insulin resistance, and NAFLD (3, 26, 35-38); b), it reduces the risk of CVD through the inhibition of vascular ectopic calcification (3, 39). This factor is reduced in individuals with acute myocardial infarction (40). Low levels of this hepatokine are correlated to higher mortality risk in a patient with coronary artery diseases (CAD) (34) and the severity of calcification (41-43). In contrast with these results, studies have shown that high levels of this hepatokine were correlated to the CVD conditions (44-46). Therefore, the association between this hepatokine and CVD is more controversial.

In 2013, Zhao et al. (46) evaluated fetuin-A serum levels in CAD patients with T2DM. They showed that subjects without CAD had significantly lower levels of serum fetuin-A as compared to CAD patients. In addition, Akin et al. in 2015 studied the correlation of fetuin-A with CAD (47) as well as coronary plaque burden (48). This hepatokine was significantly elevated in patients with CAD in comparison with patients without CAD (47, 48). By contrast, Chen et al. 2017 (34) indicated that a lower level of this hepatokine was related to an elevated risk of CVD

and all-cause mortality in CAD patients. Bilgir et al. (40) demonstrated that fetuin-A was lower in myocardial infarction and stable angina patients as compared to the control group. However, Naito et al. (49) showed that fetuin-A induced the expression of intercellular adhesion molecule-1 (IAM-1), monocyte chemoattractant protein-1 (MCP-1), E-selectin, and IL-6 in human umbilical vein endothelial cells *in vitro*. In addition, this glycoprotein was abundantly expressed in cultured human monocytes, macrophages, human aortic smooth muscle cells, human coronary artery smooth muscle cells, fibroblasts, restenosis lesions in rat carotid arteries, and atheromatous plaques in human coronary arteries. This hepatokine stimulated collagen-1 and -3 expression and cell proliferation in human aortic smooth muscle cells as well as macrophage foam cell formation related to scavenger receptors (SR-A and CD36), acyl-CoA: cholesterol acyltransferase-1 down-regulation, and ATP-binding cassette transporter A1 up-regulation (49).

As mentioned above, fetuin-A induces inflammatory cytokines, C-reactive protein (50) as well as insulin resistance (32) that these factors play important roles in the development of CVD (51). On the other hand, many studies have shown the reduction of this hepatokine following exercise (52, 53). Since, the exercise decrease the risk factors for CVD (54-56); lower fetuin-A levels following exercise, can confirm high levels of this hepatokine in patients with CVD.

Association of fetuin-A with non-alcoholic fatty liver disease

NAFLD is a metabolic disorder related to fat accumulation which is observed in people who have no alcohol consumption. Today, to create this disease, the two-hit theory is proposed to introduce the first factor to insulin resistance and the next factor of oxidative stress (57). Secretory hepatokines of hepatic cells have a significant effect on glucose and lipid metabolism, which fetuin-A is one of these hepatokines. As mentioned, fetuin-A plays a role in most metabolic diseases such as obesity, insulin resistance, and cardiovascular events and can recognize it as a biomarker (58). Considering the importance of this marker and opposite findings, it is necessary to study it in liver disorders.

Fetuin-A is one of the hepatokines with negative metabolism functions. The increase of fetuin-A is observed in the circulation of patients with NAFLD and a positive correlation is found between fetuin-A levels and the stage of liver fibrosis (7, 9). Fetuin-A stimulates the production of pro-inflammatory cytokines and activates inflammatory signals through activation of TLR4. Accordingly, it can lead to NAFLD (59). Fetuin-A levels in people with impaired glucose tolerance were negatively correlated with insulin sensitivity and positively correlated with liver fat (36, 60). Toxicity of fat and glucose by stimulating ERK1,2 and consequently increasing NF-KB, which results in increased fetuin-A upregulation causes hepatic steatosis (61). Fetuin-A in animal and human models plays a key role in insulin resistance (a risk factor for NAFLD) and acts as a phosphorylation inhibitor of insulin tyrosine kinase receptor in liver and skeletal muscles (62). The inhibition of this hepatokine could improve the impairment of lipid metabolism and palmitate-induced steatosis in liver cells through the AMPK-NFkB pathway (23).

In a study, Yusuf Yilmaz and et al. found that serum level of Fetuin-A significantly increases in adult patients with NAFLD compared to healthy controls and a significant correlation was observed between serum Fetuin-A and liver fibrosis score index (9). Conversely, in a study on the Chinese population by Zhengsen Cui et al., it was observed that fetuin-A in patients with NAFLD decreased when compared to healthy ones (63). To all appearances, Fetuin-A appears to be a marker for identifying different spectra of NAFLD and the link between diabetes and

NAFLD. However, given the contradictory findings of this hepatokine, it is important to study it further.

Association of fetuin-A with cancer

Since the genes of the metabolic pathways are out of regulation in cancer; it can be considered as the disease of metabolism (64-66). Numerous epidemiological studies have shown that certain types of cancers are more common in people with metabolic disorders (67). In cancers, in addition to the liver, cancer cells themselves also synthesize and release fetuin-A (10-12). Studies show that fetuin-A plays a role in enhancing cancer cell attachment (13), motility, and invasion (14, 15).

Fetuin-A induces cellular adhesion indirectly by regulating the secretion and stimulation of exosomes and their uptake by tumor cells (13, 68, 69). Fetuin-A has prometastatic activity and accelerates the process of colonization during calcium ion-dependent behavior (70). The Colonization process in organs is dedicated to an important part of cancer metastasis to yourself (71) and bone is a desirable place to attainment this process (72). Concerning the chemotaxis role for fetuin-A (14, 73), high affinity to hydroxyapatite crystals present in bone tissue (74), and protection of matrix metalloproteinases function (75); this biomolecule can be one of the important factors that make bone microenvironments tumor suitable for colonization of a variety of cancer cells. What is more, fetuin-A may be able to influential to determine the origin of metastatic traits in the latent and overt phases (71), as well as the severity and intensity of the colonization process (72). On balance, investigation of these ideas needs further research and a better understanding of these pathomechanism communications which can be promising and helpful in preventing and better targeting metastatic (especially latent) cancers.

Fetuin-A plays a significant role in the metabolic changes that affect cancer. In cancers that tumor initiation and progression are predominantly mediated by TGF- β signaling. Fetuin-A antagonizes this molecule during oncogenic transformation (76, 77). In 2010 Guillory et al. showed in mouse models the lacking of fetuin-A reduces breast tumor incidence, extends latency, and decreases attenuation of aging caused by oncogenes. These events were triggered by an increased TGF- β signaling pathway and decreased phosphatidylinositol 3-kinase/Akt signaling pathway (76). Concerning this problem, this question arises that is there a relationship between the production and accumulation of oncometabolite and fetuin-A (78-83). It may be possible to gain a deeper understanding of "how metabolic communication occurs in cancer" through this approach. Accordingly, it is also possible that modified forms of fetuin-A can be used soon for cancer diagnosis and staging (15).

Association of fetuin-A with metabolic bone diseases

In metabolic bone diseases such as osteoporosis, osteoporosis, osteomalacia, and rickets, the homeostasis and metabolism of calcium, phosphorus, and extracellular matrix are significantly affected (84-87). The rate of formation, the growth of hydroxyapatite crystals, and the degree of super-extracellular fluidization (serum) are influenced by several factors. Controlled regulation of these processes prevents ectopic mineralization or calcification in the physiological state (88, 89). Fetuin-A acts as a mineral chaperone (90) and due to the high affinity for the minerals (91), it can play a role in their muffling, buffering, and clearance (92-94).

Studies have shown that fetuin-A in this area has advantages compared to other proteins including 1) Despite its low circulating concentration, it has a more effective mineral-inhibiting capacity than other proteins, including albumin (91, 95, 96); 2) Compared to proteins such as osteopontin and dentin

matrix protein-1 (DMP1), it fails to require phosphorylation for its inhibitory activity (91, 94, 97-99) and 3) It is the only protein that is not synthesized locally in calcified tissues (100, 101). Babler et al. showed that the calcified soft tissues of mice lacking fetuin-A, after exposure to this glycoprotein, it returns to normal (102). In the light of above mentioned it follows that fetuin-A has a greater effect on plasma calcium distribution than albumin and modifies calcium metabolism-related changes (which involves the connection of three skeletal reservoir, soft tissues, and extracellular fluid) to a larger scale.

Fetuin-A also inhibits the formation and growth of nascent hydroxyapatite crystals, as a result, it acts as a systemic inhibitor of ectopic mineralization (88, 94). By the same token, fetuin-A is considered as one of the factors of the anti-mineralization networks (103). The point is that this inhibition only occurs at the level of de novo synthesis (inhibition of osteogenesis) (94) and it is in the "formation" phase of the bone remodeling cycle (104). As a matter of fact, estrogen is the major regulator of bone metabolism in men and women (105) and the amount of fetuin-A was significantly increased by estradiol (106). In this connection, Sari et al. showed that fetuin A levels are lower in postmenopausal osteoporotic patients than in controls (107). One of the estrogenic strategies to prevent osteoporosis is to increase osteoclast apoptosis by stimulating transforming growth factor- β (TGF- β) production in the "formation" phase of the bone remodeling cycle (108). This process is synchronized with the action of fetuin-A at this phase; on the other hand, it is associated with the inhibition of TGF- β production by fetuin-A (83). It transpires that estrogen chooses special molecules to execute its functions strategy that needs further investigation to understand more deeply the therapeutic goals.

Association of fetuin-A with chronic kidney disease

Vascular calcification is a noticeable trait of chronic kidney disease (CKD) and a definite risk factor for cardiovascular events (61). Fetuin-A prevents precipitation of calcium and phosphate and acts as a potent systemic calcification suppressor (109, 110). Several studies reported an association between low levels of serum fetuin-A with vascular/valvular calcifications, and increased fatality in subjects with end-stage renal diseases (ESRD) (109, 111-113). Caglar et al. (114) elucidated that in a group of nondiabetic subjects with different stages of CKD, fetuin-A levels diminished with a reduction in glomerular filtration rate (GFR). Coen et al. stated that hemodialysis patients with the lowest tertile of fetuin-A had the maximum CRP levels (115).

Nevertheless, there were contradictory results in previous studies on the relationship between fetuin A levels and kidney dysfunction (109). Mehrotra et al. (110) showed the association between high levels of serum fetuin-A and decreased renal clearance and remarkable proteinuria in diabetic subjects. In other research on peritoneal dialysis patients, higher albumin was related to higher fetuin-A levels (116). However, given the contradictory findings of this hepatokine, it is important to study mechanisms linking fetuin-A to CKD further.

Conclusion

These findings indicate that fetuin-A through various signaling pathways could be involved in the pathogenesis of metabolic diseases such as obesity, diabetes, insulin resistance, inflammation, CKD, CVD, cancer, bone metabolic diseases, and NAFLD. The study on these findings needs further research and a better understanding of these pathomechanism communications, which can be promising and helpful in preventing and better targeting metabolic disorders. It transpires that fetuin-A chooses special molecules to execute its functions

strategy that needs further investigation to understand more deeply the therapeutic goals.

Ethical disclosure

Nothing to declare.

Acknowledgement

Nothing to declare.

Author contributions

All the authors contributed equally.

Conflict of interest

No conflict of interest has been declared by the authors.

Funding/support

No support funding.

References

- Iroz A, Couty J-P, Postic C. Hepatokines: unlocking the multi-organ network in metabolic diseases. *Diabetologia*. 2015; 58(8):1699-703. doi:10.1007/s00125-015-3634-4
- Stefan N, Häring H-U. The role of hepatokines in metabolism. *Nat Rev Endocrinol*. 2013; 9(3):144-52. doi:10.1038/nrendo.2012.258
- Ix JH, Wassel CL, Kanaya AM, Vittinghoff E, Johnson KC, Koster A, et al. Fetuin-A and incident diabetes mellitus in older persons. *Jama*. 2008;300(2):182-8. doi:10.2337/diacare.29.04.06.dc05-1938
- Schultze H, Heide K, Haupt H. Charakterisierung eines niedermolekularen α 2 -Mukoids aus Humanserum. *Naturwissenschaften*. 1962(1):15. doi:10.1007/BF00632835
- Chatterjee P, Seal S, Mukherjee S, Kundu R, Mukherjee S, Ray S, et al. Adipocyte fetuin-A contributes to macrophage migration into adipose tissue and polarization of macrophages. *J Biol Chem*. 2013; 288(39):28324-30. doi:10.1074/jbc.C113.495473
- Jirak P, Stechemesser L, Moré E, Franzen M, Topf A, Mirna M, et al. Clinical implications of fetuin-A. *Adv Clin chem*. 2019; 89:79-130. doi:10.1016/bs.acc.2018.12.003
- Trepanowski JF, Mey J, Varady KA. Fetuin-A: a novel link between obesity and related complications. *Int J Obesity*. 2015; 39(5):734-41. doi:10.1038/ijo.2014.203
- Srinivas PR, Wagner AS, Reddy LV, Deutsch DD, Leon MA, Goustin AS, et al. Serum alpha 2-HS-glycoprotein is an inhibitor of the human insulin receptor at the tyrosine kinase level. *Molecular endocrinology (Baltimore, Md)*. 1993; 7(11):1445-55.
- Yilmaz Y, Yonal O, Kurt R, Ari F, Oral AY, Celikel CA, et al. Serum fetuin A/alpha2HS-glycoprotein levels in patients with non-alcoholic fatty liver disease: relation with liver fibrosis. *Ann Clin Biochem*. 2010; 47(Pt 6):549-53. doi:10.1258/acb.2010.010169
- Azuma K, Serada S, Takamatsu S, Terao N, Takeishi S, Kamada Y, et al. Identification of sialylated glycoproteins in Doxorubicin-treated hepatoma cells with glycoproteomic analyses. *J Proteome Res*. 2014; 13(11):4869-77. doi:10.1021/pr5004399
- Gyorffy B, Surowiak P, Budczies J, Lanczky A. Online survival analysis software to assess the prognostic value of biomarkers using transcriptomic data in non-small-cell lung cancer. *PloS one*. 2013; 8(12):e82241. doi:10.1371/journal.pone.0082241
- Szasz AM, Lanczky A, Nagy A, Forster S, Hark K, Green JE, et al. Cross-validation of survival associated biomarkers in gastric cancer using transcriptomic data of 1,065 patients. *Oncotarget*. 2016; 7(31):49322-33. doi:10.18632/oncotarget.10337
- Watson K, Koumangoye R, Thompson P, Sakwe AM, Patel T, Pratap S, et al. Fetuin-A triggers the secretion of a novel set of exosomes in detached tumor cells that mediate their adhesion and spreading. *FEBS Lett*. 2012; 586(19):3458-63. doi:10.1016/j.febslet.2012.07.071
- Nangami GN, Sakwe AM, Izban MG, Rana T, Lammers PE, Thomas P, et al. Fetuin-A (alpha 2HS glycoprotein) modulates growth, motility, invasion, and senescence in high-grade astrocytomas. *Cancer Med*. 2016; 5(12):3532-43. doi:10.1002/cam4.940
- Ochieng J, Nangami G, Sakwe A, Moyo C, Alvarez J, Whalen D, et al. Impact of Fetuin-A (AHSG) on tumor progression and type 2 diabetes. *Int J Mol Sci*. 2018;19(8):2211. doi:10.3390/ijms19082211
- Bourebaba L, Marycz K. Pathophysiological Implication of Fetuin-A Glycoprotein in the Development of Metabolic Disorders: A Concise Review. *J Clin Med*. 2019; 8(12):2033. doi:10.3390/jcm8122033
- Pérez-Sotelo D, Roca-Rivada A, Larrosa-García M, Castela C, Baamonde I, Baltar J, et al. Visceral and subcutaneous adipose tissue express and secrete functional alpha2hsglycoprotein (fetuin a) especially in obesity. *Endocrine*. 2017; 55(2):435-46. doi:10.1007/s12020-016-1132-1
- Lin X, Braymer HD, Bray GA, York DA. Differential expression of insulin receptor tyrosine kinase inhibitor (fetuin) gene in a model of diet-induced obesity. *Life Sci*. 1998; 63(2):145-53. doi:10.1016/S0024-3205(98)00250-1
- Samocha-Bonet D, Tam CS, Campbell LV, Heilbronn LK. Raised circulating fetuin-a after 28-day overfeeding in healthy humans. *Diabetes Care*. 2014; 37(1):e15-6. doi:10.2337/dc13-1728
- Brix JM, Stingl H, Höllner F, Scherthner GH, Kopp H-P, Scherthner G. Elevated Fetuin-A concentrations in morbid obesity decrease after dramatic weight loss. *J Clin Endocrinol Metabol*. 2010; 95(11):4877-81. doi:10.1210/jc.2010-0148
- Mathews ST, Singh GP, Ranalletta M, Cintron VJ, Qiang X, Goustin AS, et al. Improved insulin sensitivity and resistance to weight gain in mice null for the Ahsg gene. *Diabetes*. 2002; 51(8):2450-8. doi:10.2337/diabetes.51.8.2450
- Dasgupta S, Bhattacharya S, Biswas A, Majumdar SS, Mukhopadhyay S, Ray S, et al. NF-kappaB mediates lipid-induced fetuin-A expression in hepatocytes that impairs adipocyte function effecting insulin resistance. *Biochem J*. 2010; 429(3):451-62. doi:10.1042/BJ20100330
- Jung TW, Youn B-S, Choi HY, Lee SY, Hong HC, Yang SJ, et al. Salsalate and adiponectin ameliorate hepatic steatosis by inhibition of the hepatokine fetuin-A. *Biochem Pharmacol*. 2013; 86(7):960-9. doi:10.1016/j.bcp.2013.07.034
- Weyer C, Funahashi T, Tanaka S, Hotta K, Matsuzawa Y, Pratley RE, et al. Hypoadiponectinemia in obesity and type 2 diabetes: close association with insulin resistance and hyperinsulinemia. *J Clin Endocrinol Metabol*. 2001; 86(5):1930-5. doi:10.1210/jcem.86.5.7463
- Chatterjee P, Seal S, Mukherjee S, Kundu R, Mukherjee S, Ray S, et al. Adipocyte fetuin-A contributes to macrophage migration into adipose tissue and polarization of macrophages. *J Biol Chem*. 2013; 288(39):28324-30. doi:10.1074/jbc.C113.495473
- Stefan N, Fritsche A, Weikert C, Boeing H, Joost H-G, Häring H-U, et al. Plasma fetuin-A levels and the risk of type 2 diabetes. *Diabetes*. 2008; 57(10):2762-7. doi:10.2337/db08-0538

27. Afrisham R, Paknejad M, Soliemanifar O, Sadegh-Nejadi S, Meshkani R, Ashtary-Larky D. The influence of psychological stress on the initiation and progression of diabetes and cancer. *Int J Endocrinol Metab.* 2019; 17(2):e67400. doi:10.5812/ijem.67400
28. Sun Q, Cornelis MC, Manson JE, Hu FB. Plasma levels of fetuin-A and hepatic enzymes and risk of type 2 diabetes in women in the US. *Diabetes.* 2013; 62(1):49-55. doi:10.2337/db12-0372
29. Ix JH, Biggs ML, Mukamal KJ, Kizer JR, Ziemann SJ, Siscovick DS, et al. Association of fetuin-a with incident diabetes mellitus in community-living older adults: the cardiovascular health study. *Circulation.* 2012; 125(19):2316-22.
30. Vionnet N, Dupont S, Gallina S, Francke S, Dotte S, De Matos F, et al. Genomewide search for type 2 diabetes-susceptibility genes in French Whites: evidence for a novel susceptibility locus for early-onset diabetes on chromosome 3q27-qter and independent replication of a type 2-diabetes locus on chromosome 1q21-q24. *Am J Hum Genet.* 2000; 67(6):1470-80. doi:10.1086/316887
31. Goustin AS, Derar N, Abou-Samra AB. Ahsg-fetuin blocks the metabolic arm of insulin action through its interaction with the 95-kD β -subunit of the insulin receptor. *Cell Signal.* 2013; 25(4):981-8. doi:10.1016/j.cellsig.2012.12.011
32. Pal D, Dasgupta S, Kundu R, Maitra S, Das G, Mukhopadhyay S, et al. Fetuin-A acts as an endogenous ligand of TLR4 to promote lipid-induced insulin resistance. *Nat Med.* 2012; 18(8):1279-85. doi:10.1038/nm.2851
33. Hennige AM, Staiger H, Wicke C, Machicao F, Fritsche A, Häring H-U, et al. Fetuin-A induces cytokine expression and suppresses adiponectin production. *PLoS one.* 2008; 3(3):e1765. doi:10.1371/journal.pone.0001765
34. Chen X, Zhang Y, Chen Q, Li Q, Li Y, Ling W. Lower plasma fetuin-A levels are associated with a higher mortality risk in patients with coronary artery disease. *Arterioscler Thromb Vasc Biol.* 2017; 37(11):2213-9.
35. Nascimbeni F, Romagnoli D, Ballestri S, Baldelli E, Lugari S, Sirotti V, et al. Do nonalcoholic fatty liver disease and fetuin-A play different roles in symptomatic coronary artery disease and peripheral arterial disease?. *Diseases.* 2018;6(1):17. doi:10.3390/diseases6010017
36. Stefan N, Hennige AM, Staiger H, Machann J, Schick F, Kröber SM, et al. α 2-Heremans-Schmid glycoprotein/fetuin-A is associated with insulin resistance and fat accumulation in the liver in humans. *Diabetes Care.* 2006; 29(4):853-7. doi:10.2337/diacare.29.04.06.dc05-1938
37. Ix JH, Shlipak MG, Brandenburg VM, Ali S, Ketteler M, Whooley MA. Association between human fetuin-A and the metabolic syndrome: data from the Heart and Soul Study. *Circulation.* 2006; 113(14):1760-7. doi:10.1161/CIRCULATIONAHA.105.588723
38. Reinehr T, Roth CL. Fetuin-A and its relation to metabolic syndrome and fatty liver disease in obese children before and after weight loss. *J Clin Endocrinol Metab.* 2008; 93(11):4479-85. doi:10.1210/jc.2008-1505
39. Li W, Zhu S, Li J, Huang Y, Rongrong Z, Fan X, et al. A hepatic protein, fetuin-A, occupies a protective role in lethal systemic inflammation. *PLoS one.* 2011; 6(2):e16945. doi:10.1371/journal.pone.0016945
40. Bilgir O, Kebapcilar L, Bilgir F, Bozkaya G, Yildiz Y, Pinar P, et al. Decreased serum fetuin-A levels are associated with coronary artery diseases. *Intern Med.* 2010; 49(13):1281-5. doi:10.2169/internalmedicine.49.3223
41. Mori K, Ikari Y, Jono S, Emoto M, Shioi A, Koyama H, et al. Fetuin-A is associated with calcified coronary artery disease. *Coron Artery Dis.* 2010; 21(5):281-5. doi:10.1097/MCA.0b013e3283232fe5d5
42. Ix JH, Barrett-Connor E, Wassel CL, Cummins K, Bergstrom J, Daniels LB, et al. The associations of fetuin-A with subclinical cardiovascular disease in community-dwelling persons: the Rancho Bernardo Study. *J Am Coll Cardiol.* 2011; 58(23):2372-9.
43. Ix JH, Katz R, de Boer IH, Kestenbaum BR, Peralta CA, Jenny NS, et al. Fetuin-A is inversely associated with coronary artery calcification in community-living persons: the Multi-Ethnic Study of Atherosclerosis. *Clin Chem.* 2012; 58(5):887-95. doi:10.1373/clinchem.2011.177725
44. Weikert C, Stefan N, Schulze MB, Pischon T, Berger K, Joost H-G, et al. Plasma fetuin-a levels and the risk of myocardial infarction and ischemic stroke. *Circulation.* 2008; 118(24):2555-62.
45. Mori K, Emoto M, Araki T, Yokoyama H, Teramura M, Lee E, et al. Association of serum fetuin-A with carotid arterial stiffness. *Clin Endocrinol.* 2007; 66(2):246-50. doi:10.1111/j.1365-2265.2006.02716.x
46. Zhao Z-W, Lin C-G, Wu L-Z, Luo Y-K, Fan L, Dong X-f, et al. Serum fetuin-A levels are associated with the presence and severity of coronary artery disease in patients with type 2 diabetes. *J Biomark.* 2013; 18(2):160-4. doi:10.3109/1354750X.2012.762806
47. Akin F, Celik O, Altun I, Ayca B, Diker VO, Satilmis S, et al. Relationship of fibroblast growth factor 23 and fetuin-A to coronary atherosclerosis. *J Diabetes Complications.* 2015; 29(4):550-5. doi:10.1016/j.jdiacomp.2015.02.013
48. Akin F, Celik O, Ayca B, Altun I, Diker VO, Bıyık I, et al. Associations of fibroblast growth factor 23 and fetuin-A with coronary plaque burden and plaque composition in young adults. *J Invest Med.* 2015; 63(4):613-9. doi:10.1097/JIM.0000000000000153
49. Naito C, Hashimoto M, Watanabe K, Shirai R, Takahashi Y, Kojima M, et al. Facilitatory effects of fetuin-A on atherosclerosis. *Atherosclerosis.* 2016; 246:344-51. doi:10.1016/j.atherosclerosis.2016.01.037
50. Rittig K, Thamer C, Haupt A, Machann J, Peter A, Balletshofer B, et al. High plasma fetuin-A is associated with increased carotid intima-media thickness in a middle-aged population. *Atherosclerosis.* 2009; 207(2):341-2. doi:10.1016/j.atherosclerosis.2009.05.018
51. Ormazabal V, Nair S, Elfeky O, Aguayo C, Salomon C, Zuñiga FA. Association between insulin resistance and the development of cardiovascular disease. *Cardiovasc Diabetol.* 2018; 17(1):122. doi:10.1186/s12933-018-0762-4
52. Malin SK, Del Rincon JP, Huang H, Kirwan JP. Exercise-induced lowering of fetuin-A may increase hepatic insulin sensitivity. *Med Sci Sports Exerc.* 2014; 46(11):2085. doi:10.1249/MSS.0000000000000338
53. Yang SJ, Hong HC, Choi HY, Yoo HJ, Cho GJ, Hwang TG, et al. Effects of a three-month combined exercise programme on fibroblast growth factor 21 and fetuin-A levels and arterial stiffness in obese women. *Clin Endocrinol.* 2011;75(4):464-9. doi:10.1111/j.1365-2265.2011.04078.x
54. Mukhopadhyay S, Mondal S, Kumar M, Dutta D. Proinflammatory and antiinflammatory attributes of fetuin-a: a novel hepatokine modulating cardiovascular and glycemic outcomes in metabolic syndrome. *Endocr Pract.* 2014; 20(12):1345-51.
55. Gielen S, Laughlin MH, O'Conner C, Duncker DJ. Exercise training in patients with heart disease: review of beneficial effects and clinical recommendations. *Prog Cardiovasc Dis.* 2015; 57(4):347-55. doi:10.1016/j.pcad.2014.10.001
56. Kadoglou NP, Kottas G, Lampropoulos S, Vitta I, Liapis CD. Serum levels of fetuin-A, osteoprotegerin and osteopontin in patients with coronary artery disease: effects of statin

- (HMGCoA-reductase inhibitor) therapy. *Clin Drug Investig.* 2014; 34(3):165-71. doi:10.1007/s40261-013-0157-y
57. Peverill W, Powell LW, Skoien R. Evolving concepts in the pathogenesis of NASH: beyond steatosis and inflammation. *Int J Mol Sci.* 2014; 15(5):8591-638. doi:10.3390/ijms15058591
58. Meex RC, Watt MJ. Hepatokines: linking nonalcoholic fatty liver disease and insulin resistance. *Nat Rev Endocrinol.* 2017; 13(9):509-20. doi:10.1038/nrendo.2017.56
59. Mukhopadhyay S, Bhattacharya S. Plasma fetuin-A triggers inflammatory changes in macrophages and adipocytes by acting as an adaptor protein between NEFA and TLR-4. *Diabetologia.* 2016; 59(4):859-60. doi:10.1007/s00125-016-3866-y
60. Aroner SA, Mukamal KJ, St-Jules DE, Budoff MJ, Katz R, Criqui MH, et al. Fetuin-a and risk of diabetes independent of liver fat content: the multi-ethnic study of atherosclerosis. *Am J Epidemiol.* 2017; 185(1):54-64. doi:10.1093/aje/kww095
61. Trepanowski J, Mey J, Varady K. Fetuin-A: a novel link between obesity and related complications. *Int J Obes.* 2015; 39(5):734-41. doi:10.1038/ijo.2014.203
62. Auberger P, Falquerho L, Contreras JO, Pages G, Le Cam G, Rossi B, et al. Characterization of a natural inhibitor of the insulin receptor tyrosine kinase: cDNA cloning, purification, and anti-mitogenic activity. *Cell.* 1989; 58(4):631-40. doi:10.1016/0092-8674(89)90098-6
63. Cui Z, Xuan R, Yang Y. Serum fetuin A level is associated with nonalcoholic fatty liver disease in Chinese population. *Oncotarget.* 2017; 8(63):107149-56. doi:10.18632/oncotarget.22361
64. Collier HA. Is cancer a metabolic disease?. *Am J Pathol.* 2014; 184(1):4-17. doi:10.1016/j.ajpath.2013.07.035
65. Linehan WM, Srinivasan R, Schmidt LS. The genetic basis of kidney cancer: a metabolic disease. *Nat Rev Urol.* 2010; 7(5):277-85. doi:10.1038/nrurol.2010.47
66. Seyfried TN, Flores RE, Poff AM, D'Agostino DP. Cancer as a metabolic disease: implications for novel therapeutics. *Carcinogenesis.* 2013; 35(3):515-27. doi:10.1093/carcin/bgt480
67. Faulds MH, Dahlman-Wright K. Metabolic diseases and cancer risk. *Curr Opin Oncol.* 2012; 24(1):58-61. doi:10.1097/CCO.0b013e32834e0582
68. Kamekar S, LeBleu VS, Sugimoto H, Yang S, Ruivo CF, Melo SA, et al. Exosomes facilitate therapeutic targeting of oncogenic KRAS in pancreatic cancer. *Nature.* 2017; 546(7659):498-503. doi:10.1038/nature22341
69. Tkach M, Thery C. Communication by Extracellular Vesicles: Where We Are and Where We Need to Go. *Cell.* 2016; 164(6):1226-32. doi:10.1016/j.cell.2016.01.043
70. Kundranda MN, Henderson M, Carter KJ, Gorden L, Binhazim A, Ray S, et al. The serum glycoprotein fetuin-A promotes Lewis lung carcinoma tumorigenesis via adhesive-dependent and adhesive-independent mechanisms. *Cancer Res.* 2005; 65(2):499-506.
71. Massagué J, Obenauf AC. Metastatic colonization by circulating tumour cells. *Nature.* 2016; 529(7586):298-306. doi:10.1038/nature17038
72. Esposito M, Guise T, Kang Y. The Biology of Bone Metastasis. *Cold Spring Harb perspect Med.* 2018;8(6):a031252. doi:10.1101/cshperspect.a031252
73. Malone JD, Richards M. α 2HS glycoprotein is chemotactic for mononuclear phagocytes. *J cell Physiol.* 1987; 132(1):118-24. doi:10.1002/jcp.1041320116
74. Xie J, Baumann MJ, McCabe LR. Adsorption of serum fetuin to hydroxylapatite does not contribute to osteoblast phenotype modifications. *J Biomed Mater Res.* 2005; 73(1):39-47. doi:10.1002/jbm.a.30246
75. Ray S, Lukyanov P, Ochieng J. Members of the cystatin superfamily interact with MMP-9 and protect it from autolytic degradation without affecting its gelatinolytic activities. *Biochim Biophys Acta Proteins Proteom.* 2003; 1652(2):91-102. doi:10.1016/j.bbapap.2003.08.004
76. Guillory B, Sakwe AM, Saria M, Thompson P, Adhiambo C, Koumangoye R, et al. Lack of fetuin-A (α 2-HS-glycoprotein) reduces mammary tumor incidence and prolongs tumor latency via the transforming growth factor- β signaling pathway in a mouse model of breast cancer. *Am J Pathol.* 2010; 177(5):2635-44. doi:10.2353/ajpath.2010.100177
77. Swallow CJ, Partridge EA, Macmillan JC, Tajirian T, DiGuglielmo GM, Hay K, et al. α 2HS-glycoprotein, an antagonist of transforming growth factor β in vivo, inhibits intestinal tumor progression. *Cancer Res.* 2004; 64(18):6402-9. doi:10.1158/0008-5472.CAN-04-1117
78. Collins RR, Patel K, Putnam WC, Kapur P, Rakheja D. Oncometabolites: a new paradigm for oncology, metabolism, and the clinical laboratory. *Clin Chem.* 2017; 63(12):1812-20. doi:10.1373/clinchem.2016.267666
79. Dando I, Pozza ED, Ambrosini G, Torrens-Mas M, Butera G, Mullapilly N, et al. Oncometabolites in cancer aggressiveness and tumour repopulation. *Biol Rev.* 2019; 94(4):1530-46. doi:10.1111/brv.12513
80. Sciacovelli M, Frezza C. Oncometabolites: Unconventional triggers of oncogenic signalling cascades. *Free Radic Biol Med.* 2016; 100:175-81. doi:10.1016/j.freeradbiomed.2016.04.025
81. Wishart DS. Is cancer a genetic disease or a metabolic disease?. *EBioMedicine.* 2015; 2(6):478-9. doi:10.1016/j.ebiom.2015.05.022
82. Yang M, Soga T, Pollard PJ. Oncometabolites: linking altered metabolism with cancer. *J Clin Investig.* 2013; 123(9):3652-8. doi:10.1172/JCI67228
83. Zhou Z, Ibekwe E, Chormenkyy Y. Metabolic alterations in cancer cells and the emerging role of oncometabolites as drivers of neoplastic change. *Antioxidants.* 2018; 7(1):16. doi:10.3390/antiox7010016
84. El Demellawy D, Davila J, Shaw A, Nasr Y. Brief Review on Metabolic Bone Disease. *Acad Forensic Pathol.* 2018; 8(3):611-40. doi:10.1177/1925362118797737
85. Mankin HJ, Mankin CJ. Metabolic bone disease: a review and update. *Instr Course lect.* 2008; 57:575-93. PMID:18399611
86. Peters J, Robertson A, Godavitarne C, Rogers B. Metabolic bone disease. *Orthop Trauma.* 2017; 31(5):306-11. doi:10.1016/j.mporth.2017.07.008
87. Raubenheimer E. Part II: Metabolic bone disease: Recent developments in the pathogenesis of rickets, osteomalacia and age-related osteoporosis. *S Afr Orthop J.* 2009; 8(1):38-42.
88. Cai MM, Smith ER, Holt SG. The role of fetuin-A in mineral trafficking and deposition. *Bonekey Rep.* 2015; 4:672. doi:10.1038/bonekey.2015.39
89. Reynolds JL, Skepper JN, McNair R, Kasama T, Gupta K, Weissberg PL, et al. Multifunctional roles for serum protein fetuin-a in inhibition of human vascular smooth muscle cell calcification. *J Am Soc Nephrol.* 2005; 16(10):2920-30. doi:10.1681/ASN.2004100895
90. Jahnen-Dechent W, Schäfer C, Ketteler M, McKee MD. Mineral chaperones: a role for fetuin-A and osteopontin in the inhibition and regression of pathologic calcification. *J Mol Med.* 2008; 86(4):379-89. doi:10.1007/s00109-007-0294-y
91. Heiss A, DuChesne A, Denecke B, Grötzinger J, Yamamoto K, Renné T, et al. Structural basis of calcification inhibition by α 2-HS glycoprotein/fetuin-A formation of colloidal calciprotein particles. *J Biol Chem.* 2003; 278(15):13333-41. doi:10.1074/jbc.M210868200
92. Herrmann M, Schäfer C, Heiss A, Gräber S, Kinkeldey A, Büscher A, et al. Clearance of fetuin-A-containing calciprotein particles is mediated by scavenger receptor-A. *Circ Res.* 2012; 111(5):575-84.

93. Holt SG, Smith ER. Fetuin-A-containing calciprotein particles in mineral trafficking and vascular disease. *Nephrol Dial Transplant*. 2016; 31(10):1583-7. doi:10.1093/ndt/gfw048
94. Schinke T, Amendt C, Trindl A, Pöschke O, Müller-Esterl W, Jahn-Dechent W. The serum protein α 2-HS glycoprotein/fetuin inhibits apatite formation in vitro and in mineralizing calvaria cells a possible role in mineralization and calcium homeOSTASIS. *J Biol Chem*. 1996; 271(34):20789-96. doi:10.1074/jbc.271.34.20789
95. Schäfer C, Heiss A, Schwarz A, Westenfeld R, Ketteler M, Floege J, et al. The serum protein α 2-Heremans-Schmid glycoprotein/ fetuin-A is a systemically acting inhibitor of ectopic calcification. *J Clin Invest*. 2003; 112(3):357-66. doi:10.1172/JCI17202
96. Westenfeld R, Schäfer C, Krüger T, Haarmann C, Schurgers LJ, Reutelingsperger C, et al. Fetuin-A protects against atherosclerotic calcification in CKD. *J Am Soc Nephrol*. 2009; 20(6):1264-74. doi:10.1681/ASN.2008060572
97. Deshpande AS, Fang P-A, Zhang X, Jayaraman T, Sfeir C, Beniash E. Primary structure and phosphorylation of dentin matrix protein 1 (DMP1) and dentin phosphophoryn (DPP) uniquely determine their role in biomineralization. *Biomacromolecules*. 2011; 12(8):2933-45. doi:10.1021/bm2005214
98. Hunter GK. Role of osteopontin in modulation of hydroxyapatite formation. *Calcif Tissue Int*. 2013; 93(4):348-54. doi:10.1007/s00223-013-9698-6
99. Smith ER, Ford ML, Tomlinson LA, Rajkumar C, McMahon LP, Holt SG. Phosphorylated fetuin-A-containing calciprotein particles are associated with aortic stiffness and a procalcific milieu in patients with pre-dialysis CKD. *Nephrol Dial Transplant*. 2012; 27(5):1957-66. doi:10.1093/ndt/gfr609
100. Saunders NR, Deal A, Dziegielewska KM, Reader M, Sheardown S, Møllgård K. Expression and distribution of fetuin in the developing sheep fetus. *Histochemistry*. 1994; 102(6):457-75. doi:10.1007/BF00269578
101. Terkelsen OB, Jahn-Dechent W, Nielsen H, Moos T, Fink E, Nawratil P, et al. Rat fetuin: distribution of protein and mRNA in embryonic and neonatal rat tissues. *Anat Embryol*. 1998; 197(2):125-33.
102. Babler A, Schmitz C, Buescher A, Herrmann M, Gremse F, Gorgels TG, et al. Soft tissue calcification in mice is governed by fetuin-A, pyrophosphate and magnesium. *BioRxiv*. 2019:577239. doi:10.1101/577239
103. Li Q, Uitto J. Mineralization/anti-mineralization networks in the skin and vascular connective tissues. *Am J Pathol*. 2013; 183(1):10-8. doi:10.1016/j.ajpath.2013.03.002
104. Binkert C, Demetriou M, Sukhu B, Szwercas M, Tenenbaum HC, Dennis JW. Regulation of osteogenesis by fetuin. *J Biol Chem*. 1999; 274(40):28514-20. doi:10.1074/jbc.274.40.28514
105. Khosla S, Monroe DG. Regulation of bone metabolism by sex steroids. *Cold Spring Harb perspect Med*. 2018; 8(1):a031211. doi:10.1101/cshperspect.a031211
106. Kim SW, Choi J-W, Lee DS, Yun JW. Sex hormones regulate hepatic fetuin expression in male and female rats. *Cell Physiol Biochem*. 2014; 34(2):554-64. doi:10.1159/000363022
107. Sari A, Uslu T. The relationship between fetuin-a and bone mineral density in postmenopausal osteoporosis/Postmenapozal Osteoporozda Fetuin-A ve Kemik Mineral Yögunlugu Arasindaki Iliski. *Turk J Rheumatol*. 2013; 28(3):195. doi:10.5606/tjr.2013.2468
108. Gao Y, Qian W-P, Dark K, Toraldo G, Lin AS, Guldberg RE, et al. Estrogen prevents bone loss through transforming growth factor β signaling in T cells. *Proc Nat Acad Sci*. 2004; 101(47):16618-23.
109. Mutluay R, DEĞERTEKİN CK, Derici Ü, Gültekin S, Gönen S, Arinsoy ST, et al. Serum fetuin-A is associated with the components of MIAC (malnutrition, inflammation, atherosclerosis, calcification) syndrome in different stages. *Turk J Med Sci*. 2019; 49(1):327-35.
110. Mehrotra R, Westenfeld R, Christenson P, Budoff M, Ipp E, Takasu J, et al. Serum fetuin-A in nondialyzed patients with diabetic nephropathy: relationship with coronary artery calcification. *Kidney Int*. 2005; 67(3):1070-7. doi:10.1038/sj.ki.5002178
111. Stenvinkel P, Wang K, Qureshi AR, Axelsson J, Pecoits-Filho R, Gao P, et al. Low fetuin-A levels are associated with cardiovascular death: impact of variations in the gene encoding fetuin. *Kidney Int*. 2005; 67(6):2383-92. doi:10.1111/j.1523-1755.2005.00345.x
112. Hermans M, Brandenburg V, Ketteler M, Kooman J, Van der Sande F, Boeschoten E, et al. Association of serum fetuin-A levels with mortality in dialysis patients. *Kidney Int*. 2007; 72(2):202-7. doi:10.1038/sj.ki.5002178
113. Metry G, Stenvinkel P, Qureshi A, Carrero J, Yilmaz M, Barany P, et al. Low serum fetuin-A concentration predicts poor outcome only in the presence of inflammation in prevalent haemodialysis patients. *Eur J Clin Invest*. 2008; 38(11):804-11. doi:10.1111/j.1365-2362.2008.02032.x
114. Caglar K, Yilmaz MI, Saglam M, Cakir E, Kilic S, Sonmez A, et al. Serum fetuin-a concentration and endothelial dysfunction in chronic kidney disease. *Nephron Clin Pract*. 2008; 108(3):c233-c40. doi:10.1159/000120209
115. Coen G, Manni M, Agnoli A, Balducci A, Dessi M, De Angelis S, et al. Cardiac calcifications: fetuin-A and other risk factors in hemodialysis patients. *Asaio J*. 2006; 52(2):150-6. doi:10.1097/01.mat.0000202606.44826.6b
116. Wang AY-M, Woo J, Lam CW-K, Wang M, Chan IH-S, Gao P, et al. Associations of serum fetuin-A with malnutrition, inflammation, atherosclerosis and valvular calcification syndrome and outcome in peritoneal dialysis patients. *Nephrol Dial Transplant*. 2005; 20(8):1676-85. doi:10.1093/ndt/gfh891